

PAPER • OPEN ACCESS

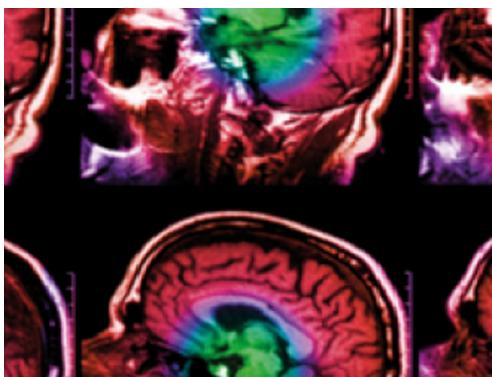
VHEE beam dosimetry at CERN Linear Electron Accelerator for Research under ultra-high dose rate conditions

To cite this article: Daniela Poppinga *et al* 2021 *Biomed. Phys. Eng. Express* **7** 015012

View the [article online](#) for updates and enhancements.

You may also like

- [Review on the characteristics of radiation detectors for dosimetry and imaging](#)
Joao Seco, Ben Clasie and Mike Partridge
- [Advances in kilovoltage x-ray beam dosimetry](#)
Robin Hill, Brendan Healy, Lois Holloway et al.
- [A new approach to radiochromic film dosimetry based on non-local means](#)
César Rodríguez, Alfonso López-Fernández and Diego García-Pinto



IPEM | IOP

Series in Physics and Engineering in Medicine and Biology

Your publishing choice in medical physics,
biomedical engineering and related subjects.

Start exploring the collection—download the
first chapter of every title for free.

**OPEN ACCESS****PAPER**

VHEE beam dosimetry at CERN Linear Electron Accelerator for Research under ultra-high dose rate conditions

RECEIVED
22 July 2020

REVISED
30 October 2020

ACCEPTED FOR PUBLICATION
16 November 2020

PUBLISHED
3 December 2020

Original content from this work may be used under the terms of the Creative Commons Attribution 4.0 licence.

Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI.



Daniela Poppinga¹ , Rafael Kranzer^{1,5}, Wilfrid Farabolini^{3,4}, Antonio Gilardi^{2,4,6} , Roberto Corsini⁴, Vanessa Wyrwoll⁴, Hui Khee Looe⁵, Björn Delfs⁵, Lukas Gabrisch⁵ and Björn Poppe⁵

¹ PTW Freiburg, Freiburg, Germany

² Federico II, DIETI, University of Napoli, Napoli, Italy

³ CEA Saclay, IRFU-DACM, France

⁴ CERN, CH1211 Geneva, Switzerland

⁵ University Clinic for Medical Radiation Physics, Medical Campus Pius Hospital, Carl von Ossietzky University, Oldenburg, Germany

⁶ National Institute for Nuclear Physics (INFN), Section of Napoli, Italy

E-mail: Daniela.poppinga@ptwdosimetry.com

Keywords: FLASH, dosimetry, recombination loss

Abstract

The aim of this work is the dosimetric characterization of a plane parallel ionization chamber under defined beam setups at the CERN Linear Electron Accelerator for Research (CLEAR). A laser driven electron beam with energy of 200 MeV at two different field sizes of approximately 3.5 mm FWHM and approximately 7 mm FWHM were used at different pulse structures. Thereby the dose-per-pulse range varied between approximately 0.2 and 12 Gy per pulse. This range represents approximately conventional dose rate range beam conditions up to ultra-high dose rate (UHDR) beam conditions. The experiment was based on a water phantom which was integrated into the horizontal beamline and radiochromic films and an Advanced Markus ionization chamber was positioned in the water phantom. In addition, the experimental setup were modelled in the Monte Carlo simulation environment FLUKA. In a first step the radiochromic film measurements were used to verify the beamline setup. Depth dose distributions and dose profiles measured by radiochromic film were compared with Monte Carlo simulations to verify the experimental conditions. Second, the radiochromic films were used for reference dosimetry to characterize the ionization chamber. In particular, polarity effects and the ion collection efficiency of the ionization chamber were investigated for both field sizes and the complete dose rate range. As a result of the study, significant polarity effects and recombination loss of the ionization chamber were shown and characterized. However, the work shows that the behavior of the ionization chamber at the laser driven beam line at the CLEAR facility is comparable to classical high dose-per-pulse electron beams. This allows the use of ionization chambers on the CLEAR system and thus enables active dose measurement during the experiment. Compared to passive dose measurement with film, this is an important step forward in the experimental equipment of the facility.

1. Introduction

Over the past several years, the question of whether ultra-high-dose rate (UHDR) beams might offer a new modality for cancer treatment is one of the most discussed subjects in modern radiotherapy. In 2014 Favaudon *et al* (Favaudon *et al* 2014) realized that mice lung tissue showed a significantly improved radiation tolerance, when irradiated with electrons with nominal dose rates of about 100 Gy s^{-1} . In recent years

several studies have confirmed, discussed and modelled the effects in other animal and cell models (Loo *et al* 2017, Montay-Gruel *et al* 2017, Buonanno *et al* 2019, Pratx and Kapp 2019a, 2019b, Voznin *et al* 2019b, 2019a). These dose rates result in treatment times of less than a second hence the name FLASH became common for this new technique. Radiobiologically it is not yet fully understood why and how the FLASH normal tissue-sparing effect occurs. At the moment the hypothesis favored by most groups is a

local oxygen depletion caused by the interacting beam, thus reducing the oxygen-stabilisation of the radiation damages (Durante *et al* 2018, Pratx and Kapp 2019b). However studies not reporting a clear FLASH-effect have also been published, although a very high dose rate (100 Gy s⁻¹ protons) was applied (Beyreuther *et al* 2019). Analyzing the differences it became obvious that the used particles (e.g. electron, protons or photons) the microscopic structure of the beam may play an important role: Accelerators deliver the nominal dose in pulsed beams with repetition rates in the ms-range and pulse widths in the μ s-range (Bruggmoser *et al* 2007, Lang *et al* 2012, Durante *et al* 2018). Often the single pulse is sub-divided into even shorter ns or even ps-bunches (in this case a pulse is often called 'train'). When using this beam structure, the dose within a single pulse or train can reach instantaneous dose rates in the range of 10¹⁰ Gy s⁻¹. Thus, irradiation of a nominal dose rate in the order of several 100 Gy s⁻¹ can be achieved by many combinations of pulse or train structures with varying repetition frequencies, energies and particles. Having all these degrees of freedom in mind, it seems not surprising that the FLASH-effect can only be found under special combinations of these beam parameters. It is therefore one of the most important questions (not only for the clinical transfer) to evaluate the beam parameters which show the FLASH-effect (Durante *et al* 2018, Harrington 2019).

To realize a teletherapy approach, electron energies of more than 100 MeV are necessary to deliver the desired dose to the needed depths in the body. Therefore, different groups have investigated the possibility of radiation therapy with very high electron beams (VHEE) and published theoretical planning studies with VHEE (Bazalova-Carter *et al* 2015, Subiel *et al* 2017) as well as first dosimetric characterizations (Subiel *et al* 2014).

The characteristics of dosimeters under UHDR conditions must be fully understood to achieve accurate dosimetry in these challenging conditions. It must be expected that several detectors either saturate in these UHDR conditions or show a significant dose-rate-dependence in terms of recombination loss in comparison to conventional dose rates. Several studies have started to investigate this issue (Petersson *et al* 2017, Jorge *et al* 2019). For the Advanced Markus ionization chamber, Petersson *et al* (Petersson *et al* 2017) showed recombination losses up to 70% under typical UHDR conditions.

A promising accelerator system for a systematic analysis of the detector behavior in UHDR conditions using electron beams of several hundred MeV is the CERN Linear Electron Accelerator for Research (CLEAR) (Corsini *et al* 2018, Gamba *et al* 2018, Sjobak *et al* 2019). It provides a laser driven high energy electron beam with a wide range of possible beam characteristics. It is possible to generate a beam that approximates a conventional irradiation condition

with dose rate similar to that used currently in radiation therapy although there are differences in the microscopic structure. In the same setup the beam can be changed to UHDR conditions. To achieve this, the number of bunches as well as the charge per pulse can be varied within a large range. With this flexibility and the option to reach clinically relevant electron beams, the CLEAR facility offers the opportunity to explore UHDR beams.

The aim of this work is the characterization of detector behavior in the defined beam setups at the CLEAR facility. For this study an electron beam with 200 MeV at two different field sizes of approximately 3.5 mm FWHM and 7 mm FWHM were used. For both field sizes, different pulse structures were investigated which resemble UHDR conditions. Additionally, one pulse was defined to reproduce a conventional dose rate electron beam as closely as possible. Based on film dosimetry as reference, this work develops a procedure for practical dosimetry at the facility.

2. Materials and methods

2.1. Beam configuration

The experiment was performed at the TeraHertz (THz) test stand at the CERN Linear Electron Accelerator for Research (CLEAR) (Gamba *et al* 2018) facility at CERN (Switzerland). This accelerator complex is capable of providing electrons with energies from 55 MeV up to 200 MeV. The beam is generated using a photocathode in Cs₂Te then thanks to three accelerating stage, powered by two RF sources, it is possible to reach the top energy of 200 MeV. The accelerator line continues with a diagnostic section, where different setups allow a precise bunch length measurement and beam energy measurement (Arpaia *et al* 2020). Following, two irradiation areas are located, the first is VESPER, a test stand for irradiation installed on a spectrometer line, and the second is the so called 'in air' TeraHertz (THz) test-stand (Lagzda *et al* 2020, McManus *et al* 2020). Between the VESPER and the THz test-stands, several experiments are installed, from study related to X-band accelerating structure (Arpaia *et al* 2019) to plasma-based focusing lens (Lindstrøm *et al* 2018).

The beam size at the phantom entrance is extracted from the beam transverse profile revealed by a Yttrium-Aluminum-Garnet (YAG) screen of 0.5 mm thickness. The picture acquired by a monochromatic digital camera is calibrated using a calibration pattern fixed on the screen and taking into account the tilt angle of the screen. The beam charge was measured using an Integrating Current Transformer (ICT), model ICT-055-5.0 (Bergoz Instrumentation, France) installed just after the exit window and 50 cm upstream to the phantom entrance, with its electronic unit BCM-IHR-E.

For the experiment four different beam time structures with different number of bunches per pulse

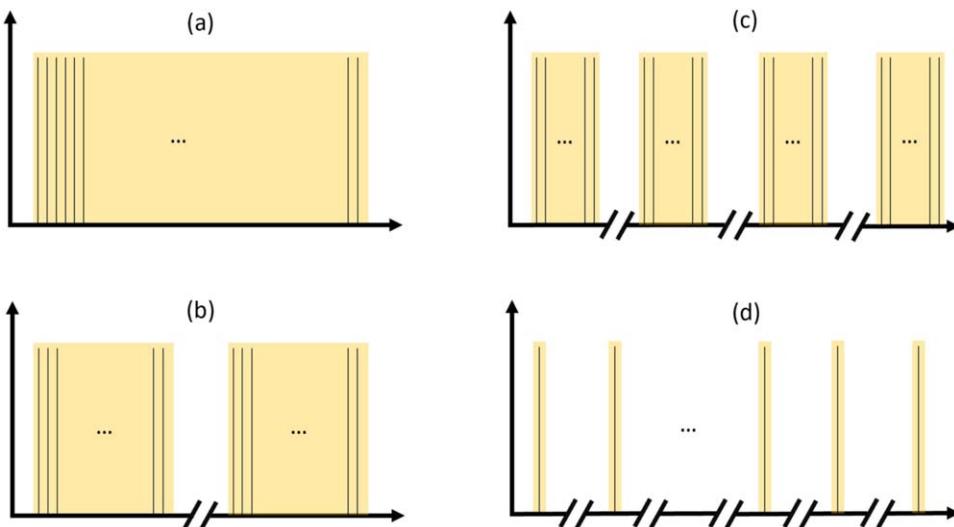


Figure 1. Beam time structures: (a) one pulse with 100 bunches ('very high') (b) two pulses with 50 bunches per pulse ('high') (c) 4 pulses with 25 bunches per pulse ('medium') (d) 1–2 bunches per pulse and so many pulses that a total charge of 2 nC was achieved ('low'). The black lines symbolize the individual bunches, the yellow area symbolizes a pulse structure. Time between two bunches was 666 ps (1.5 GHz) and time between two pulses was 1.2 s.

Table 1. Overview of the beam time structure parameters.

	VERY HIGH	HIGH	MEDIUM	LOW
Number of pulses	1	2	4	36–80
Time length of one pulse	66 ns	33 ns	16 ns	~10 ps
Time between two pulses			1.2 s	
Number of bunches per pulse	100	50	25	1–2
Time between two bunches			666 ps (1.5 GHz)	

and different numbers of pulse per measurement were used. Thus a large range of different dose per pulse values could be realized. Each bunch had a charge of approximately 20 pC and a time length of a few ps, whereas the time length between two bunches was approximately 666 ps (1.5 GHz). The four conditions are sketched in figure 1 and described in the following: (i) Very High: 1 pulse with 100 bunches, which corresponds to a charge of approximately 2 nC and pulse length of 66 ns. (ii) High: 2 pulses with 50 bunches per pulse, which corresponds to a total charge of approximately 2 nC and a length per pulse of 33 ns. (iii) Medium: 4 pulses with 25 bunches per pulse, which corresponds to a total charge of approximately 2 nC and length per pulse of 16 ns. (iv) Low: 1–2 bunches per pulse and so many pulses that a total charge of approximately 2 nC was achieved. Time between two pulses was 1.2 s for all beam time structures. An overview of all parameters is given in table 1.

During the irradiation, in addition to the four different beam time structures, two different transverse beam sizes were used (approximately 3.5 and 7 mm FWHM at the phantom entrance). The beam size of 3.5 mm FWHM is obtained using the various set of quadrupoles of the CLEAR beam line. To enlarge it to 7.0 mm, a scattering foil (silicon, 0.5 mm thick

approximately) is inserted just upstream to the exit window (0.1 mm aluminum), the quadrupole current settings remaining unchanged.

2.2. Phantom setup and ionization chamber measurement

A water phantom with the dimensions of $30 \times 30 \times 10$ cm³ was positioned on a motorized table. The phantom includes a probe holder for biological experiments with 8 Eppendorf tubes. The phantom was placed in the horizontal beam line as shown in figure 2. For this experiment, film measurements were performed at three positions: in front of the phantom, in front of the probe holder as well as at the back of the probe holder. In addition, an Advanced Markus ionization chamber type 34045 (PTW Freiburg, Germany) was placed independently from the water phantom movement at 72 mm water depth in the phantom. The chamber was connected using 42 m cable to an UNIDOSwebline (PTW Freiburg, Germany) electrometer with a high voltage of 400 V.

The absorbed dose to water measured by the ionization chamber can be calculated according to international protocols like IAEA TRS 398 (Andreo *et al* 2006) according to equation (1)

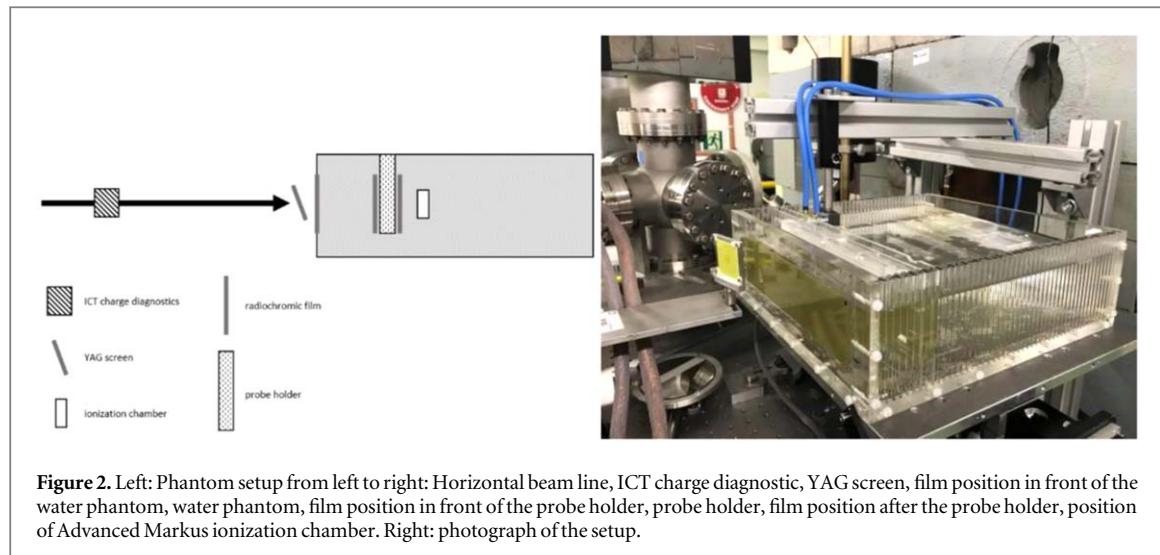


Figure 2. Left: Phantom setup from left to right: Horizontal beam line, ICT charge diagnostic, YAG screen, film position in front of the water phantom, water phantom, film position in front of the probe holder, probe holder, film position after the probe holder, position of Advanced Markus ionization chamber. Right: photograph of the setup.

$$\text{Dose to water} = M \cdot N \cdot k_{TP} \cdot k_E \cdot k_P \cdot k_S \quad (1)$$

where M is the measured signal, the calibration factor N for irradiation with Co60 source, temperature and pressure correction in terms of k_{TP} , the beam quality correction k_E , the polarity effect correction k_P as well as the correction for recombination loss k_S . Under Co60 irradiation with $+400\text{V}$, the calibration factor N for the used Advanced Markus chamber was $N = 1.601 \cdot 10^9 \text{ Gy C}^{-1}$. Water temperature was 19° and air pressure was 960 hPa which leads to a correction of $k_{TP} = 1.05$. The beam quality correction k_E is not available in standard protocols for a 200 MeV electron beam. Therefore, the correction was simulated as described in section D.2 and determined to be $k_E = 0.79$.

In contrast to the small magnitude of the polarity effect at standard reference conditions described in international protocols, the polarity effect under these conditions is expected to be quite high (Petersson *et al* 2017). For this reason, the polarity effect was analyzed in detail. For this purpose, the ionization chamber was irradiated with approximately 2 nC beam charge with different number of pulses for both beam sizes at $+400 \text{ V}$ and -400 V . For each measurement, the ICT charge was recorded.

Each data set was fitted to a power model as follows:

$$M_{\text{voltage, beam size}}(\text{trains}) = a \cdot \text{trains}^b + c \quad (2)$$

where M describes the chamber measurement normalized by the recorded ICT charge value. According to AAPM TG 51 (Almond *et al* 1999), the polarity correction factor k_P was calculated as follows:

$$k_{P,+400\text{V}} = \left| \frac{M_{+400\text{V}} + M_{-400\text{V}}}{2M_{+400\text{V}}} \right| \quad (3a)$$

$$k_{P,-400\text{V}} = \left| \frac{M_{+400\text{V}} + M_{-400\text{V}}}{2M_{-400\text{V}}} \right| \quad (3b)$$

where $k_{P,+400\text{V}}$ describes the correction factor for positive voltage and $k_{P,-400\text{V}}$ for negative voltage and $M_{+/-400\text{V}}$ are defined by the fit functions according to equation (2).

2.3. Film measurement

For all measurements, EBT3 films with batch number 10231801 were used.

2.3.1. Film calibration

Considering the original film orientation, 15 film pieces of $3.5 \text{ cm} \times 5.0 \text{ cm}$ size were cut and irradiated with a conventional radiation therapy linear accelerator (Siemens Primus, Siemens, Germany) at an electron beam of 21 MeV under known dosimetry conditions. The 15 film pieces were irradiated with dose values between 0.1 Gy and 20.4 Gy. As shown by Subiel *et al* (Subiel *et al* 2014) and Jaccard *et al* (Jaccard *et al* 2017b) an energy independence of EBT2 and EBT3 films exists to very high electron energies. Therefore a calibration at 21 MeV was assumed to be appropriate.

The film pieces were scanned with Epson 10000XL scanner and transparency unit using EpsonScan software with all auto corrections turned off. All scans were performed with 300 dpi and the films were scanned in landscape orientation. The scanned images were analyzed with Matlab 2019a and imaging processing toolbox. Since the film irradiations in the center of the field exceeded dose values of 8 Gy, the green color channel information was evaluated (Devic *et al* 2009, Borca *et al* 2013), for which a calibration curve has been obtained as shown in figure 3. The calibration curve was fitted by an exponential function:

$$\begin{aligned} \text{Dose}(\text{PixelValue}) = & -0.000\ 1586 \cdot \exp \\ & \times (0.000\ 1891 \cdot \text{pixelValue}) + 70.69 \\ & \cdot \exp(-0.000\ 1231 \cdot \text{pixelValue}) \end{aligned} \quad (4)$$

2.3.2. Film measurement—depth dose distribution

Again, by considering the original film orientation, film strips of $25.4 \text{ cm} \times 5.1 \text{ cm}$ size were cut. On a PMMA holder two film strips were stacked and fixed in the phantom parallel to the beam orientation using the laser alignment system as shown in figure 4. The

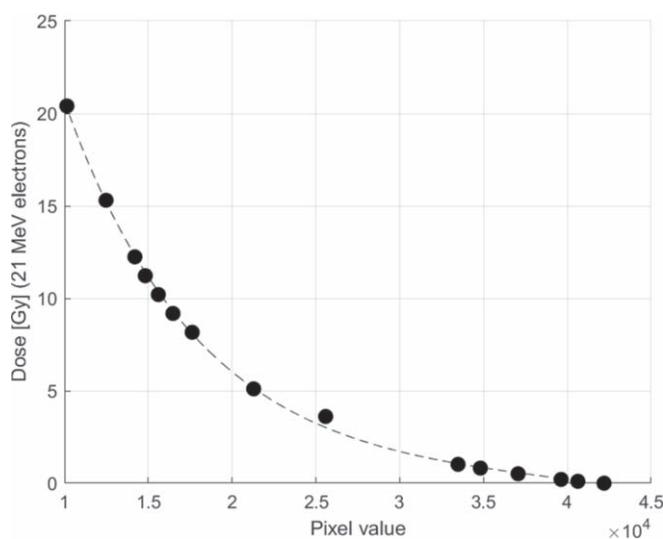


Figure 3. Calibration function of EBT3 films irradiated with 21 MeV electron beam.

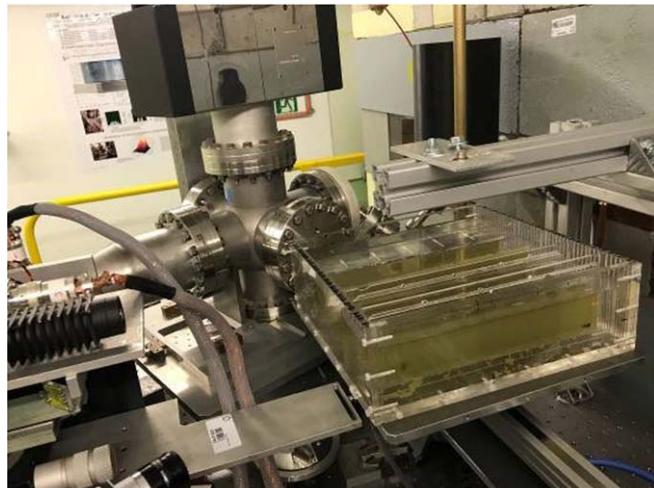


Figure 4. Phantom setup for depth dose measurement using EBT3 film. The films were positioned in water parallel to beam direction as shown by the arrow.

films were positioned directly after the phantom wall, so that the measurement starts at depth = (15 ± 5) mm. In the literature (Aldelaijan *et al* 2010, León-Marroquín *et al* 2018) it was shown that films absorb water at the edges if staying in water for extended time, which could affect film response. To minimize this effect, the edges were taped as they were submerged in the water phantom overnight due to radiation protection related access restrictions. Since no more than six film strips could be positioned in the phantom simultaneously, the beam time structures ‘very high’, ‘high’ and ‘low’ were irradiated each with two films. The beam time structure ‘medium’ was not irradiated.

2.3.3. Film measurement—absorbed dose

The films were prepared as for the depth dose distribution measurements. For each measurement, a film strip was positioned perpendicular to the beam

direction in front and another at the back of the probe holder, corresponding to depths of 38 mm and 48 mm, respectively, as shown in figure 2. Figure 5 shows an example irradiated film strip. Since the beam has to be checked first after every change of beam parameters, a part of the film was strongly blackened by these test measurements. Therefore, only positions #0–#5 were analyzed.

The same scan protocols as for the calibration films were used. The calibration curve was applied to the film strips to obtain the absolute dose. For each measurement the beam spot was separated, and the center position of the spot was analyzed. Then, the dose at the center position was calculated as well as the dose averaged in a circle of 5 mm diameter around the center position (corresponding to 2734 pixels). The averaging area of 5 mm diameter corresponds to the diameter of the sensitive volume of the ionization

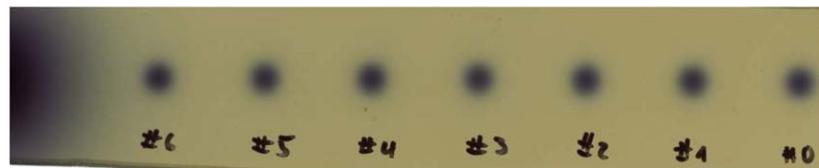


Figure 5. EBT3 film used for absolute dose measurement. Example scanned image. Position #0–#5 were analyzed.

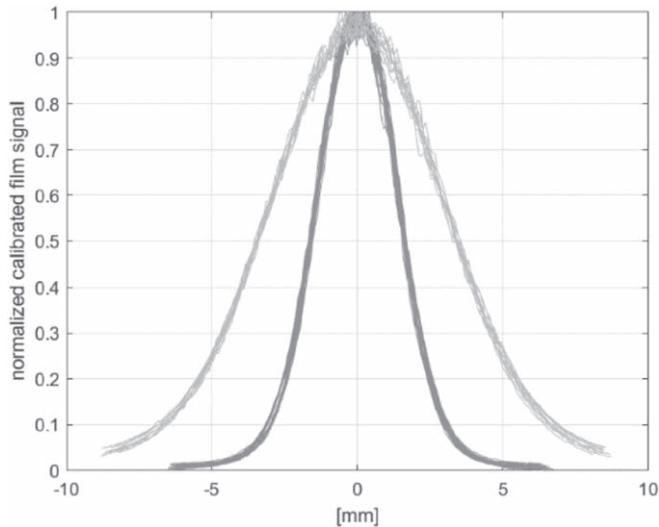


Figure 6. Beam profiles of all film measurements positioned in front of the water phantom.

chamber and takes into account the volume averaging effect of the ionization chamber. An overall uncertainty of 5% is assumed for the entire film evaluation. The calibration curve was used for both irradiation conditions (parallel and perpendicular, see Arjomandy *et al* 2012).

2.4. Monte Carlo simulation

Part of the Monte Carlo simulations were performed using FLUKA Monte Carlo simulation package. FLUKA version 2011.3 and the graphical user interface FLAIR were used (Vlachoudis 2009, Böhlen *et al* 2014, Battistoni *et al* 2015). No variance reduction technique was applied. For further simulations of the beam quality correction EGSnrc was used applying the user code egs-chamber. Range rejection and Russian Roulette were used for variance reduction.

2.4.1. Depth dose distribution

The setup was based on a cylindrical geometry consisting of water with radius of 10 cm and length of 30 cm. A monoenergetic electron beam with energy of 200 MeV and a Gaussian beam shape was used. The FWHM of the beam was simulated in one case with 3.23 mm and in the other case with 6.88 mm according to the results from the film measurements as shown in figure 6.

2.4.2. Beam quality correction k_E

For the simulation of the correction factor k_E , the absorbed dose to water was simulated with Co60 source as well as with monoenergetic electron beam with 200 MeV. The correction factor k_E is defined as:

$$k_E = \frac{(s_{w,a}^\Delta)_{200\text{MeV},7.2\text{cm}} \cdot p_{200\text{MeV},7.2\text{cm}}}{(s_{w,a}^\Delta)_{\text{Co60},5\text{cm}} \cdot p_{\text{Co60},5\text{cm}}} \quad (5)$$

The first term in both the numerator and denominator is chamber independent, where $(s_{w,a}^\Delta)_{\text{Co60},5\text{cm}}$ defines the stopping power ratio between air and water at Co60 irradiation and $(s_{w,a}^\Delta)_{200\text{MeV},7.2\text{cm}}$ the stopping power ratio under 200 MeV electron beam irradiation. The second term describes the fluence perturbation p considering the fluence perturbation of non-ideal cavities caused by real ionization chambers.

The Advanced Markus chamber was modelled according to manufacturer's specifications. Cutoff energies were defined at 0.521 MeV for electrons and 0.01 MeV for photons. The simulation was performed under reference conditions using a 10 cm \times 10 cm field size as well as for Gaussian shape beams used in this study. In each simulation, the deposited dose in the chamber's sensitive volume was scored. For validation, the value of k_E was simulated for 6 MeV electron beam and compared to the value given in international dosimetry protocol (IAEA TRS 398).

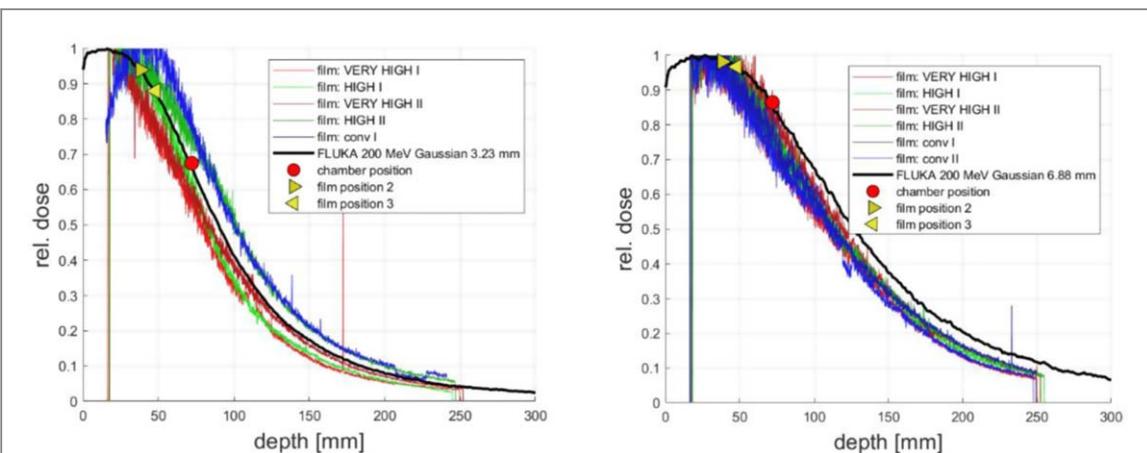


Figure 7. Depth dose distribution measured with EBT3 film compared with Monte Carlo simulated depth dose distribution. Left: beam configuration with 3.5 mm FWHM Right: beam configuration with 7 mm FWHM. For both configurations the film and chamber positions are marked.

3. Results

3.1. Beam characterization

3.1.1. Beam size

Based on film strips positioned in front of the water phantom free in air, the beam size was analyzed. For each spot, the film data was calibrated, and the profiles were centered and normalized. All analyzed beam profiles are shown in figure 6. For narrower transverse beam size the mean FWHM value is (3.23 ± 0.09) mm, for the scattered beam a FWHM of (6.88 ± 0.11) mm was determined. No differences in the beam transverse profiles between different beam time structures are observed.

3.1.2. Depth dose distribution

The depth dose distributions measured by EBT3 film and the simulated depth dose distribution for all beam configurations are shown in figure 7. Due to alignment errors, the second film for the 'LOW' beam time structure for the beam size of 3.5 mm field could not be analyzed. No difference in the depth dose distributions between the different beam time structures for 3.5 mm and 7 mm beam size can be observed within the measurement accuracy of the film measurement. Furthermore, there is a good agreement with the Monte Carlo simulation within the measurement accuracy. Accuracy limitation is mainly due to the uncertainty of the film positioning in the water phantom; the films alignment along the beam axis being critical, especially for narrow beam size and at the entrance of the phantom.

To compensate for the different positions of films and ionization chamber measurements, corrections derived from Monte Carlo calculated depth dose distributions were applied. These measurement positions are marked in figure 7. The corresponding corrections required between the films and chamber positions are 0.72 (film position 2) and 0.77 (film position 3) for the 3.5 mm transverse beam size and 0.88 (film position 2)

and 0.89 (film position 3) for the 7 mm beam size. These correction factors are not depending on the beam time structures.

3.2. Chamber measurement

Based on the Monte Carlo simulation of the 200 MeV electron beam, an energy correction factor $k_E = (0.79 \pm 0.005)$ has been obtained for both the beam sizes with FWHM 3.5 and 7 mm. The results of the polarity correction are shown in figure 8. It shows the fitting functions of the measurement data according to equation (2) with 68% uncertainty level ($k = 1$). With decreasing number of pulses the chamber signal relative to the ICT value also decreases. This is caused by a collection efficiency loss of the ionization chamber. However, this decreasing of the chamber signal is polarity dependent. As shown in figure 8, the chamber signal relative to ICT is higher for negative polarity than for positive polarity. The difference between the polarities increases with decreasing number of pulses. Figure 8 shows the polarity correction factor for both beam sizes for positive chamber voltage exemplary. The polarity correction factor for positive polarity computed according to equation (3) is up to (1.39 ± 0.06) for beam sizes 7.0 mm FWHM and 1 pulse per measurement. The correction factors are summarized in table 2.

The ionization chamber measurements are presented in table 2 for different beam conditions. The column $M \cdot N \cdot k_{TP} \cdot k_E \cdot k_P$ represents the dose value before the application of the correction factor for the recombination loss k_s . The ion collection efficiency will be derived from the comparison between the film and ionization chamber measurements that is described in the next section.

3.3. Ion collection efficiency of ionization chamber

As mentioned above the ion collection efficiency of ionization chamber measurements UHDR conditions is a key parameter for precise dose to water determination. As listed in table 2, film measurements were

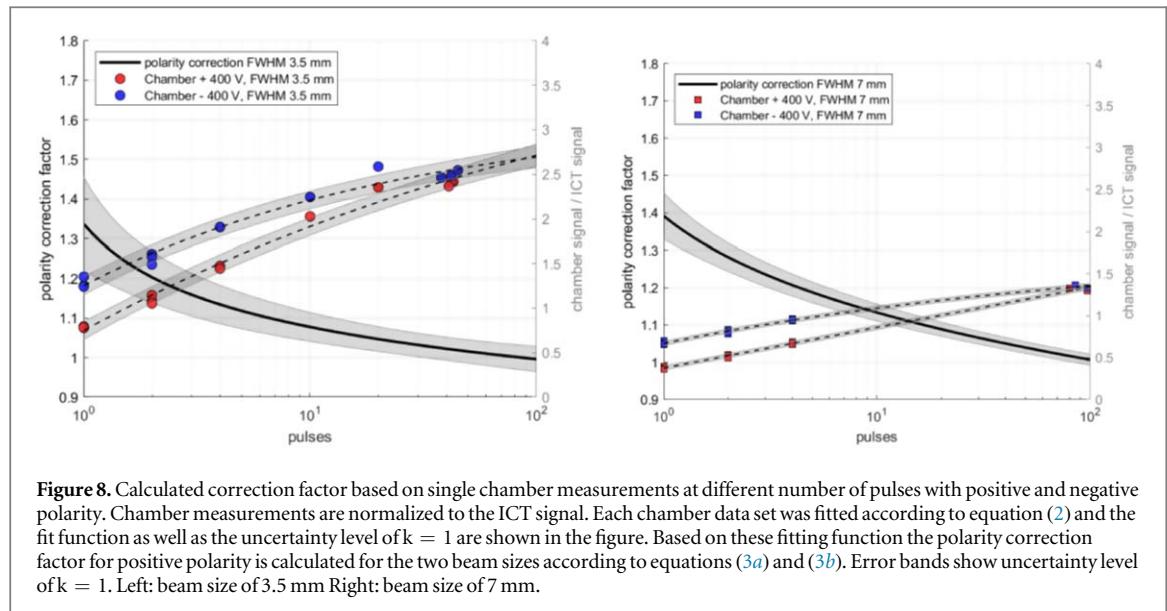


Table 2. Summary of film and detector measurements. Beam size: small or broad beam size; beam structure according to figure 1; number of pulses; Dose per pulse: calculated based on the depth corrected mean value of the 5 mm averaged film data. Film measurement: 5 mm average film value normalized to ICT value and depth corrected (uncertainty 5%), Ionization chamber measurement: calculation according to equation (2) before the application of recombination loss correction factor ($M \cdot N \cdot k_{TP} \cdot k_E \cdot k_P$), normalized to ICT value.

Beam size	Beam configuration	No of pulses	Dose per pulse	Film measurement front of probe holder	Film measurement back of probe holder	Chamber measurement	Error Dose Error [Gy]
				[Gy/nC]	[Gy/nC]	[Gy/nC]	[Gy]
3.5 mm	LOW	36	0.20	4.08	4.15	3.98	0.21
		37	0.20	4.10	4.07	3.85	0.21
		37	0.21	4.21	4.06	3.81	0.21
		39	0.19	3.80	4.26	3.94	0.21
		39	0.20	4.12	4.37	3.96	0.21
		41	0.17	4.18	4.00	3.76	0.19
		45	0.16	4.30	3.98	3.69	0.18
	MEDIUM	4	2.31	4.32	4.34	2.29	0.24
		4	2.17	4.42	4.30	2.37	0.23
	HIGH	2	4.76	4.25	4.41	1.80	0.25
		2	4.46	3.40	3.52	1.48	0.25
		2	4.99	4.54	4.17	1.71	0.25
		2	4.98	4.56	4.03	1.68	0.25
	VERY HIGH	1	7.82	4.53	4.50	1.62	0.33
		1	9.08	4.44	4.59	1.53	0.36
		1	9.17	4.60	4.61	1.54	0.36
		1	6.70	4.47	4.18	1.38	0.25
		1	10.15	4.57	4.34	1.42	0.38
7 mm	LOW	80	0.13	2.29	2.15	1.83	0.12
		85	0.12	2.25	2.12	1.81	0.13
	HIGH	2	5.03	2.11	1.89	0.85	0.16
		2	5.09	2.11	1.89	0.83	0.16
	VERY HIGH	1	9.60	2.23	2.02	0.66	0.18
		1	11.65	2.24	2.07	0.67	0.22

performed at two positions in the water phantom simultaneously with each ionization chamber measurement. By comparing the dose values determined by the film measurement and the dose values obtained from the ionization chamber measurements, the efficiency of the ionization chamber and thus the recombination loss of the chamber can be determined

as follows:

$$eff = \frac{M \cdot N \cdot k_{Pol} \cdot k_E \cdot k_{TP}}{Dose_{Film, \text{5mm, depth corrected}}} \quad (6)$$

Where the numerator describes the dose calculation based on the chamber measurement without concerning recombination loss as listed in table 2 and the

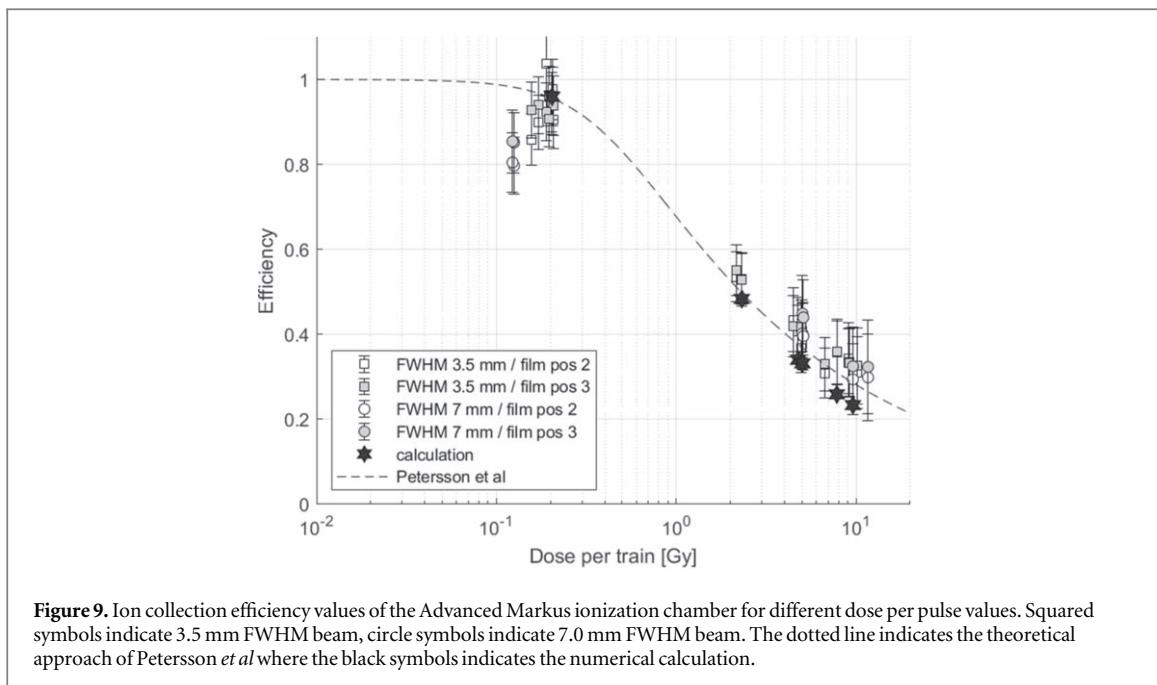


Figure 9. Ion collection efficiency values of the Advanced Markus ionization chamber for different dose per pulse values. Squared symbols indicate 3.5 mm FWHM beam, circle symbols indicate 7.0 mm FWHM beam. The dotted line indicates the theoretical approach of Petersson *et al* where the black symbols indicates the numerical calculation.

denominator describes the dose value based on the film measurement, whereas the film measurement was averaged according to the chamber size and depth corrected as listed in table 2 for both film positions.

To analyze the efficiency of the chamber, the dose per pulse value was determined. For this purpose, the depth corrected film value was divided by number of pulse per measurement and listed in table 2.

Finally, the efficiency of the Advanced Markus chamber was calculated by dividing the ionization chamber value normalized to ICT measurement through the depth corrected film value normalized to ICT measurement. The ion collection efficiency values plotted against the dose per pulse are shown in figure 9. No difference between the two beam sizes was observed. Furthermore, the results from the two film positions agree which indicate that the simulation of the depth dose curve as shown in figure 7 is valid. For the highest dose per pulse value a chamber ion collection efficiency of approximately 30% is observed.

4. Discussion

The beam structure used in this study differs significantly from conventional clinical beam structures. Linear accelerators used in conventional radiation therapy have a pulsed beam structure and energies up to 20 MeV. The pulse repetition frequency is between 50–400 Hz and the pulse lengths are in the range of a few microseconds. Typical dose per pulse values are up to few mGy per pulse. The electron beam with energy of 200 MeV used here is generated by many bunches that can be combined to form a pulse. The microstructure of the bunches is not detectable by the ionization chamber. Therefore, the pulse structure can be compared with the pulse structure.

The time structure of the beam used here has pulse lengths of up to 66 ns. Conventional linear accelerators,

on the other hand, have pulse lengths of several microseconds (Bruggmoser *et al* 2007, Kry *et al* 2012). In this experiment, beam doses between 0.1 and 12 Gy were applied per pulse. Conventional linear accelerators apply radiation doses of up to a few mGy per pulse. Petersson *et al* (Petersson *et al* 2017) have used an electron linear accelerator (Oriatron eRT6) in their study where the applied beam dose per pulse can be varied between 0.1 mGy and 10 Gy. Therefore, a detailed comparison to the study by Petersson *et al* is appropriate. However, the time structure of the beam used in Petersson *et al* with pulse lengths between 0.5 μ s and 1.8 μ s is comparable to conventional linacs and not to the time structure used here.

Radiochromic film has been used in many previous publications to determine the absorbed dose to water. The film does not seem to show any dependence of the dose rate, also for ultra-high dose rate conditions (Jaccard *et al* 2016, 2017a, Favaudon *et al* 2019). The films used in this study have been calibrated using 20 MeV electron beams, as has been also performed by Subiel *et al* (Subiel *et al* 2014) for their measurements in 165 MeV electron beam. Since a systematic study on the energy dependence of radiochromic films up to 200 MeV does not exist so far, a higher uncertainty associated with the film measurements has to be considered.

The approach used in this study based on EBT3 film to determine the dose and a simultaneous measurement with an ionization chamber which was directly comparable to the study by Petersson *et al* (Petersson *et al* 2017). Furthermore, both studies investigated the Advanced Markus chamber. Even if the beam configurations are not directly identical, figure 9 shows that the empirical model of (Petersson *et al* 2017) (Eq (10)) for pulse length of 500 ns, the smallest pulse duration investigated in their study, agree with our results. In addition, the ion collection

efficiency of the Advanced Markus chamber was calculated numerically by an approach of Gotz *et al* (Gotz *et al* 2017), which was extended and validated into the UHDR range by Kranzer *et al* (Kranzer *et al* 2020). In the numerical calculation, the actual time length of each pulse of $0.066\ \mu\text{s}$ for VERY HIGH beam time structure, $0.033\ \mu\text{s}$ for HIGH beam time structure, $0.016\ \mu\text{s}$ for MEDIUM beam time structure and $0.001\ \mu\text{s}$ for LOW beam time structure were used. Except for the data points at the lowest dose per pulse, the results of the measurement also show good agreement to the numerical calculation.

Therefore, the result in figure 9 is quite remarkable. A dependence of the time structure of the beam as well as of the electron energy is not observable within the measurement uncertainty. The result shows that it seems possible to perform reliable ionization chamber dosimetry under UHDR beam conditions. It is necessary to introduce additional procedures, most importantly to characterize and correct the saturation as shown in figure 9. It is also noteworthy that at VHEE, the water-to-air stopping power ratio, which is the detector dependent term of the beam quality correction factor, approaches a constant value. Therefore, a small spectral perturbation, such as that introduced by the scatterer to increase the beam cross section in the experiment, is not expected to cause a noticeable change in the k_E as demonstrated in the Monte Carlo results.

Another interesting result of the study is the characterization of the polarity effect of the ionization chamber. Under conventional beam conditions the polarity effect is usually very low with values below 1% as specified by IEC 60731, under special conditions the effect can also amount to values of a few percent. Under conventional beam conditions the effect is mainly caused by the fact that chamber components are irradiated, thereby providing a signal contribution in addition to signal caused by the air volume of the chamber. This additional signal contribution is independent of the chamber voltage, resulting in a polarity dependent total signal.

Under the very high dose per pulse conditions used here it can be assumed that an additional effect is involved. The dose per pulse is so high that free charge carriers are created in the air volume which lead to a temporary space charge effect within the chamber. As a result, the signal contribution from the air volume is changed. This effect is therefore proportional to the dose per pulse. Figure 8 shows a strong dependence of the dose per pulse. So, the theory seems to be possible, but should be investigated in further studies. In any case, this work shows how important it is to consider the polarity effect at UHDR.

5. Conclusion

Within the study, a procedure was developed which allows a practicable dosimetry at the CLEAR facility at the CERN. Based on film dosimetry as reference the behavior of the Advanced Markus ionization chamber at ultrahigh

dose-rates was analyzed. It shows that the behavior of the ionization chamber at the laser driven beam line at the CLEAR facility can be considered and corrected for in a way similar to very high dose per pulse electron beams. This allows the use of ionization chambers on the CLEAR system and thus enables active dose measurement during the experiment. Compared to passive dose measurement with film, this is an important step forward in the experimental equipment of the facility.

Acknowledgments

The authors gratefully acknowledge Ruben Garcia Alia for fruitful discussions and his comments on the manuscript.

This project has received funding from the EMPIR programme co-financed by the Participating States and from the European Union's Horizon 2020 research and innovation programme. The authors acknowledge the GMEE for supporting Antonio Gilardi.

Conflict of interest

Daniela Poppinga and Rafael Kranzer are employees of PTW Freiburg. The authors acknowledge the GMEE for supporting Antonio Gilardi.

ORCID iDs

Daniela Poppinga  <https://orcid.org/0000-0003-4765-3183>

Antonio Gilardi  <https://orcid.org/0000-0002-4773-5798>

References

Aldelaijan S, Devic S, Mohammed H, Tomic N, Liang L-H, DeBlois F and Seuntjens J 2010 Evaluation of EBT-2 model GAFCHROMIC™ film performance in water *Med. Phys.* **37** 3687–93

Almond P R, Biggs P J, Coursey B M, Hanson W F, Huq M S, Nath R and Rogers D W O 1999 AAPM's TG-51 protocol for clinical reference dosimetry of high-energy photon and electron beams *Med. Phys.* **26** 1847–70

Andreop P, Burns D T and Hohlfeld K 2006 IAEA TRS-398 *Absorbed Dose Determination in External Beam Radiotherapy: An International Code of Practice for Dosimetry based on Standards of Absorbed Dose to Water* Online: http://naweb.iaea.org/nahu/DMRP/documents/CoP_V12_2006-06-05.pdf

Arpaia P, Corsini R, Gilardi A and Sjobak K N 2019 Beam-based alignment of the CLIC high-gradient X-Band accelerating structure using beam-screen 2019 *IEEE Int. Instrumentation and Measurement Technology Conf. (I2MTC) 2019 (Auckland, New Zealand)* pp 1–6

Arpaia P, Corsini R, Gilardi A, Mostacci A, Sabato L and Sjobak K N 2020 Enhancing particle bunch-length measurements based on Radio Frequency Deflector by the use of focusing elements *Sci. Rep.* **10** 1–12

Arjomandy B, Tailor R, Zhao L and Devic S 2012 EBT2 film as a depth-dose measurement tool for radiotherapy beams over a wide range of energies and modalities *Med. Phys.* **39** 912–21

Battistoni G *et al* 2015 Overview of the FLUKA code *Ann. Nucl. Energy* **82** 10–8

Bazalova-Carter M, Qu B, Palma B, Härdemark B, Hynning E, Jensen C, Maxim P G and Loo B W 2015 Treatment planning for radiotherapy with very high-energy electron beams and comparison of VHEE and VMAT plans *Med. Phys.* **42** 2615–25

Beyreuther E, Brand M, Hans S, Hideghéty K, Karsch L, Leßmann E, Schürer M, Szabó E R and Pawelke J 2019 Feasibility of proton FLASH effect tested by zebrafish embryo irradiation *Radiother. Oncol.* **139** 46–50

Böhlen T T, Cerutti F, Chin M P W, Fassò A, Ferrari A, Ortega P G, Mairani A, Sala P R, Smirnov G and Vlachoudis V 2014 The FLUKA code: developments and challenges for high energy and medical applications *Nucl. Data Sheets* **120** 211–4

Borca V C, Pasquino M, Russo G, Grossi P, Cante D, Sciaci P, Girelli G, La Porta M R and Tofani S 2013 Dosimetric characterization and use of GAFCHROMIC EBT3 film for IMRT dose verification *J. Appl. Clin. Med. Phys.* **14** 158–71

Bruggmoser G, Saum R, Schmachtenberg A, Schmid F and Schüle E 2007 Determination of the recombination correction factor k_{sf} for some specific plane-parallel and cylindrical ionization chambers in pulsed photon and electron beams *Phys. Med. Biol.* **52** N35–N50

Buonanno M, Grilj V and Brenner D J 2019 Biological effects in normal cells exposed to FLASH dose rate protons *Radiother. Oncol.* **139** 51–5

Corsini R *et al* 2018 First experiments at the clear user facility *Proc. 9th Int. Part. Accel. Conf. (IPAC'18) (Vancouver, BC, Canada, Apr. 4–4066)* (<https://doi.org/10.18429/JACoW-IPAC2018-THPMF014>)

Devic S, Tomic N, Soares C G and Podgorsak E B 2009 Optimizing the dynamic range extension of a radiochromic film dosimetry system *Med. Phys.* **36** 429–37

Durante M, Bräuer-Krisch E and Hill M 2018 Faster and safer? FLASH ultra-high dose rate in radiotherapy *Br. J. Radiol.* **91** 6–9

Favaudon V *et al* 2014 Ultrahigh dose-rate FLASH irradiation increases the differential response between normal and tumor tissue in mice *Sci. Transl. Med.* **6** 1–10

Favaudon V, Lentz J-M, Heinrich S, Patriarca A, de Marzi L, Fouillade C and Dutreix M 2019 Time-resolved dosimetry of pulsed electron beams in very high dose-rate, FLASH irradiation for radiotherapy preclinical studies *Nucl. Instruments Methods Phys. Res. Sect. A Accel. Spectrometers, Detect. Assoc. Equip.* **944** 162537

Gamba D *et al* 2018 The CLEAR user facility at CERN *Nucl. Instruments Methods Phys. Res. Sect. A Accel. Spectrometers, Detect. Assoc. Equip.* **909** 480–3

Gotz M, Karsch L and Pawelke J 2017 A new model for volume recombination in plane-parallel chambers in pulsed fields of high dose-per-pulse *Phys. Med. Biol.* **62** 8634–54

Harrington K J 2019 Ultrahigh dose-rate radiotherapy: next steps for FLASH-RT *Clin. cancer Res. an Off. J. Am. Assoc. Cancer Res.* **25** 3–5

Jaccard M, Petersson K, Buchillier T, Germond J-F, Durán M T, Vozenin M-C, Bourhis J, Bochud F O and Bailat C 2016 High dose-per-pulse electron beam dosimetry: usability and dose-rate independence of EBT3 Gafchromic films *Med. Phys.* **44** 725–35

Jaccard M, Durán M T, Petersson K, Germond J-F, Liger P, Vozenin M-C, Bourhis J, Bochud F and Bailat C 2017a High dose-per-pulse electron beam dosimetry: commissioning of the Oriatron eRT6 prototype linear accelerator for preclinical use *Med. Phys.* **45** 863–74

Jaccard M, Petersson K, Buchillier T, Germond J-F, Duran M T, Vozenin M-C, Bourhis J, Bochud F O and Bailat C 2017b High dose-per-pulse electron beam dosimetry: usability and dose-rate independence of EBT3 Gafchromic films *Med. Phys.* **44** 725–35

Jorge P G *et al* 2019 Dosimetric and preparation procedures for irradiating biological models with pulsed electron beam at ultra-high dose-rate *Radiother. Oncol.* **139** 34–9

Kranzer R, Poppinga D, Weidner J, Schueller A, Hackel T, Looe H K and Poppe B 2020 Ion collection efficiency of ionization chambers in ultra-high dose-per-pulse electron beams *Med. Phys.* **47**

Kry S, Popple R, Molineu A and Followill D 2012 Ion recombination correction factors (pion) for varian truebeam high dose rate therapy beams *J. Appl. Clin. Med. Phys.* **39** 3790

Lagzda A, Angal-Kalinin D, Jones J, Aitkenhead A, Kirkby K J, MacKay R, Van Herk M, Farabolini W, Zeeshan S and Jones R M 2020 Influence of heterogeneous media on Very High Energy Electron (VHEE) dose penetration and a Monte Carlo-based comparison with existing radiotherapy modalities *Nucl. Instruments Methods Phys. Res. Sect. B Beam Interact. with Mater. Atoms* **482** 70–81

Lang S, Reggiori G, Puxeu Vaqué J, Calle C, Hrbacek J, Klöck S, Scorsetti M, Cozzi L and Mancosu P 2012 Pretreatment quality assurance of flattening filter free beams on 224 patients for intensity modulated plans: a multicentric study *Med. Phys.* **39** 1351–6

León-Marroquín E Y, Lárraga-Gutiérrez J M, Herrera-González J A, Camacho-López M A, Villarreal Barajas J E and García-Garduño O A 2018 Investigation of EBT3 radiochromic film's response to humidity *J. Appl. Clin. Med. Phys.* **19** 283–90

Lindström C A *et al* 2018 Overview of the CLEAR plasma lens experiment *Nucl. Instruments Methods Phys. Res. Sect. A Accel. Spectrometers, Detect. Assoc. Equip.* **909** 379–82

Loo B W, Schuler E, Lartey F M, Rafat M, King G J, Trovati S, Koong A C and Maxim P G 2017 (P003) delivery of ultra-rapid flash radiation therapy and demonstration of normal tissue sparing after abdominal irradiation of mice *Int. J. Radiat. Oncol.* **98** E16

McManus M, Romano F, Lee N D, Farabolini W, Gilardi A, Royle G, Palmans H and Subiel A 2020 The challenge of ionisation chamber dosimetry in ultra-short pulsed high dose-rate very high energy electron beams *Sci. Rep.* **10** 1–11

Montay-Gruel P *et al* 2017 Irradiation in a flash: unique sparing of memory in mice after whole brain irradiation with dose rates above 100 Gy s⁻¹ *Radiother. Oncol.* **124** 365–9

Petersson K, Jaccard M, Germond J-F, Buchillier T, Bochud F, Bourhis J, Vozenin M-C and Bailat C 2017 High dose-per-pulse electron beam dosimetry—a model to correct for the ion recombination in the Advanced Markus ionization chamber *Med. Phys.* **44** 1157–67

Pratx G and Kapp D S 2019a A computational model of radiolytic oxygen depletion during FLASH irradiation and its effect on the oxygen enhancement ratio *Phys. Med. Biol.* **64**

Pratx G and Kapp D S 2019b Ultra-high-dose-rate FLASH irradiation may spare hypoxic stem cell niches in normal tissues *Int. J. Radiat. Oncol. Biol. Phys.* **105** 190–2

Sjobak K N *et al* 2019 Status of the clear electron beam user facility at cern *Int. Part. Accel. Conf.* **983–6**

Subiel A *et al* 2014 Dosimetry of very high energy electrons (VHEE) for radiotherapy applications: using radiochromic film measurements and Monte Carlo simulations *Phys. Med. Biol.* **59** 5811–29

Subiel A, Moskvin V, Welsh G H, Cipiccia S, Reboreda D, DesRosiers C and Jaroszynski D A 2017 Challenges of dosimetry of ultra-short pulsed very high energy electron beams *Phys. Medica* **42** 327–31

Vlachoudis V 2009 Flair: a powerful but user friendly graphical interface for FLUKA *International Conference on Mathematics, Computational Methods and Reactor Physics* 2 790–800

Vozenin M-C, Hendry J H and Limoli C L 2019a Biological benefits of ultra-high dose rate FLASH radiotherapy: sleeping beauty awoken *Clin. Oncol.* **31** 407–15

Vozenin M C *et al* 2019b The advantage of FLASH radiotherapy confirmed in mini-pig and cat-cancer patients *Clin. Cancer Res.* **25** 35–42